

MASS SPECTROMETRIC STUDY OF CYCLODEPSIPEPTIDES

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Recent years have witnessed a heightened interest in the chemistry of depsipeptides. Unfortunately, the methods used at present for investigating this class of compounds are quite restricted and not very specific. We therefore considered it desirable to investigate the possibilities afforded by the mass spectrometric method in the solution of structuroanalytic problems of depsipeptide chemistry. We chose for the principal objects of the study a number of cyclic di-, tetra-, hexa-, octa- and dodecadepsipeptides, analogs of the enniatin and valinomycin antibiotics. Our main objective was to elucidate the dependence of the mass spectrometric behaviour of these compounds upon the size of the ring and the nature and sequence of the amino and hydroxy acid residues.

An examination of the spectra of these cyclodepsipeptides

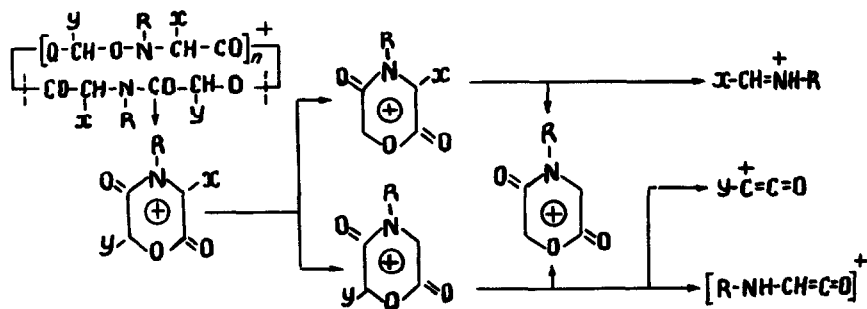
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showed that the molecular ions ( $M^+$ ) undergo three basic types of fragmentation: the morpholine type (Scheme 1), the  $CO_2$  type (Scheme 2) and the acylaminoketene type (Scheme 3). The type of fragmentation strongly depends upon the size of the ring and each group of cyclodepsipeptides displays a characteristic type of fragmentation. The morpholine type (Scheme 1) is common to all the depsipeptides investigated, but its contribution to the mass spectrum is closely related to the stability of the ring. It is weakly manifested in the case of the strained (cf. 1) 12-membered cyclotetradepsipeptides (Figs. 1,2), but it is predominant in the case of cyclooctadepsipeptides with the less strained 24-membered ring (Fig. 4) and especially in the case of cyclohexadepsipeptides with nonstrained 18-membered ring (Fig. 3). The  $CO_2$  type of fragmentation (Scheme 2), beginning with elimination of  $CO_2$ , is the most clearly expressed in the mass spectra of strained cyclotetradepsipeptides (Figs. 1,2). Finally the acylaminoketene type of fragmentation (Scheme 3) is present to about the same extent (although very little in comparison with the morpholine type) in the mass spectra of cyclohexa- and cyclooctadepsipeptides (Figs. 3,4).

The morpholine type of fragmentation begins with formation of the molecular ion of 2,5-dioxomorpholine ( $F_1$ ) the further decomposition of which is determined by the stability of the 6-membered ring (Scheme 1). The first act of fragmentation of  $F_1$  consists in elimination of olefin from the amino or hydroxy acid residue (or from both of them), the morpholine ring being retained and the fragments  $F_2$ ,  $F_3$  and  $F_5$  being formed. The next step in the fragmentation process is opening of the ring resulting in the formation of the amine fragment  $F_4$  from  $F_2$

S c h e m e 1  
Morpholine type of cyclodepsipeptide fragmentation



$n = 0, 1, 2$ ;  $R = H, Me$ ;  $X = CHMe_2, CH_2CHMe_2, CH(Me)Et$ ;  $Y = Me, CHMe_2$

T a b l e 1  
Intensities of the characteristic peaks  
of 3,6-dialkyl-2,5-dioxomorpholines<sup>\*/</sup>

	$F_1$	$F_2$	$F_3$	$F_4$	$F_5$	$F_6$	$F_7$
I	199	$\frac{2,2}{62}(157)$ <sup>**/</sup>		$\frac{16,1}{100}(72)$	$\frac{0,5}{3,1}(115)$	$\frac{2,6}{60}(83)$	$\frac{1,0}{6,2}(57)$
II	213	$\frac{10,5}{61}(171)$ <sup>**/</sup>		$\frac{1,7}{10}(86)$	$\frac{1,2}{7}(129)$	$\frac{17,3}{100}(83)$	$\frac{6,0}{35}(71)$
III	227	$\frac{4,1}{20}(185)$	$\frac{13,5}{67}(171)$	$\frac{2,7}{13,5}(100)$	$\frac{1,5}{7,5}(129)$	$\frac{20,1}{100}(83)$	$\frac{7,5}{35}(71)$
IV	227	$\frac{5,5}{43}(185)$	$\frac{0,95}{7,5}(171)$	$\frac{7,7}{61}(100)$	$\frac{8,0}{62}(129)$	$\frac{4,0}{31}(83)$	$\frac{1,3}{10}(71)$

<sup>\*/</sup>  $J$  per cent of  $\Sigma J$  per cent of  $J_{max}$  (m/e)    I  $R=H, X=Y=CHMe_2$ ; II  $R=Me, X=Y=CHMe_2$ ;  
<sup>\*\*/</sup>  $F_2+F_3$     III  $R=Me, X=CH(Me)Et, Y=CHMe_2$ ;  
 IV  $R=Me, X=CH_2CHMe_2, Y=CHMe_2$ .

and the ketene fragment  $F_6$  from  $F_3$ . Both pathways can be observed in the mass spectra of the 2,5-dioxomorpholines investigated (I-IV)(Table 1), the relative contribution of each of them

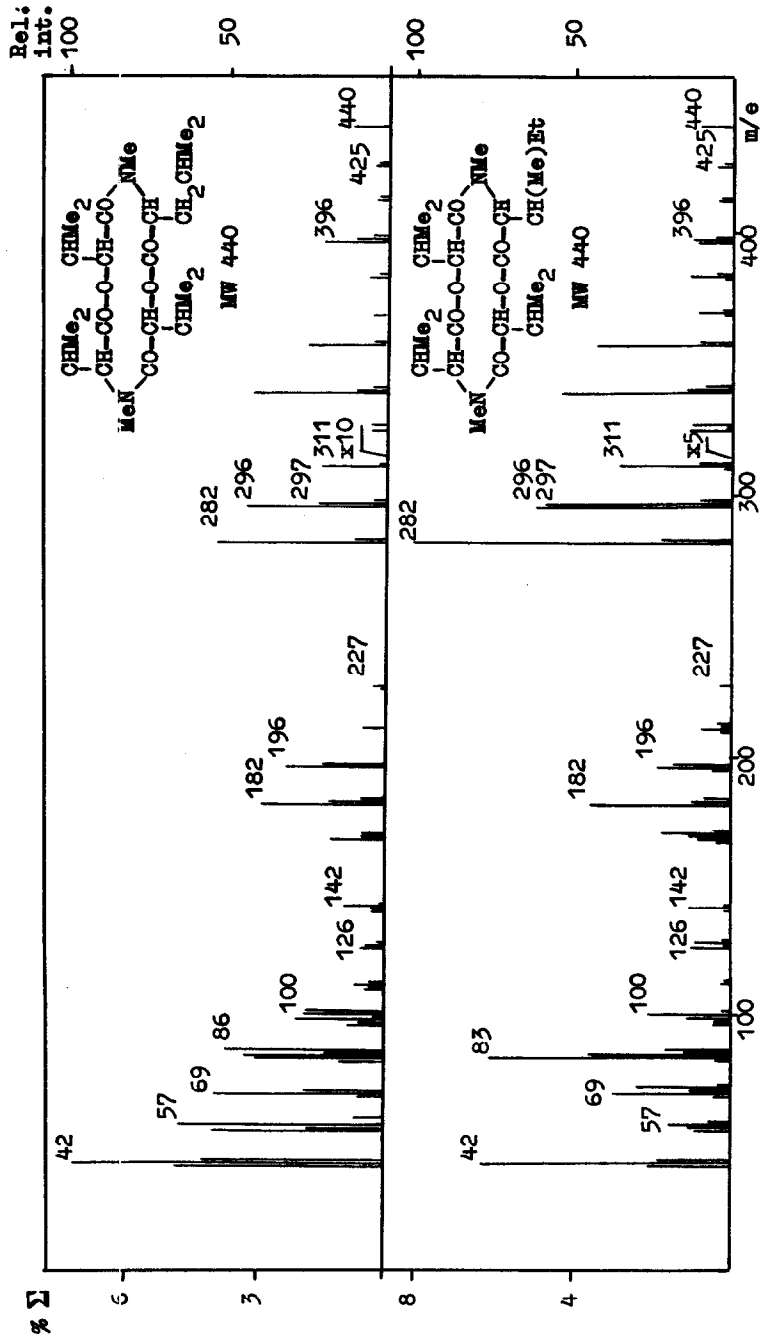


Fig. 1. Mass spectra of unsymmetrical cyclotetradepsipeptides.

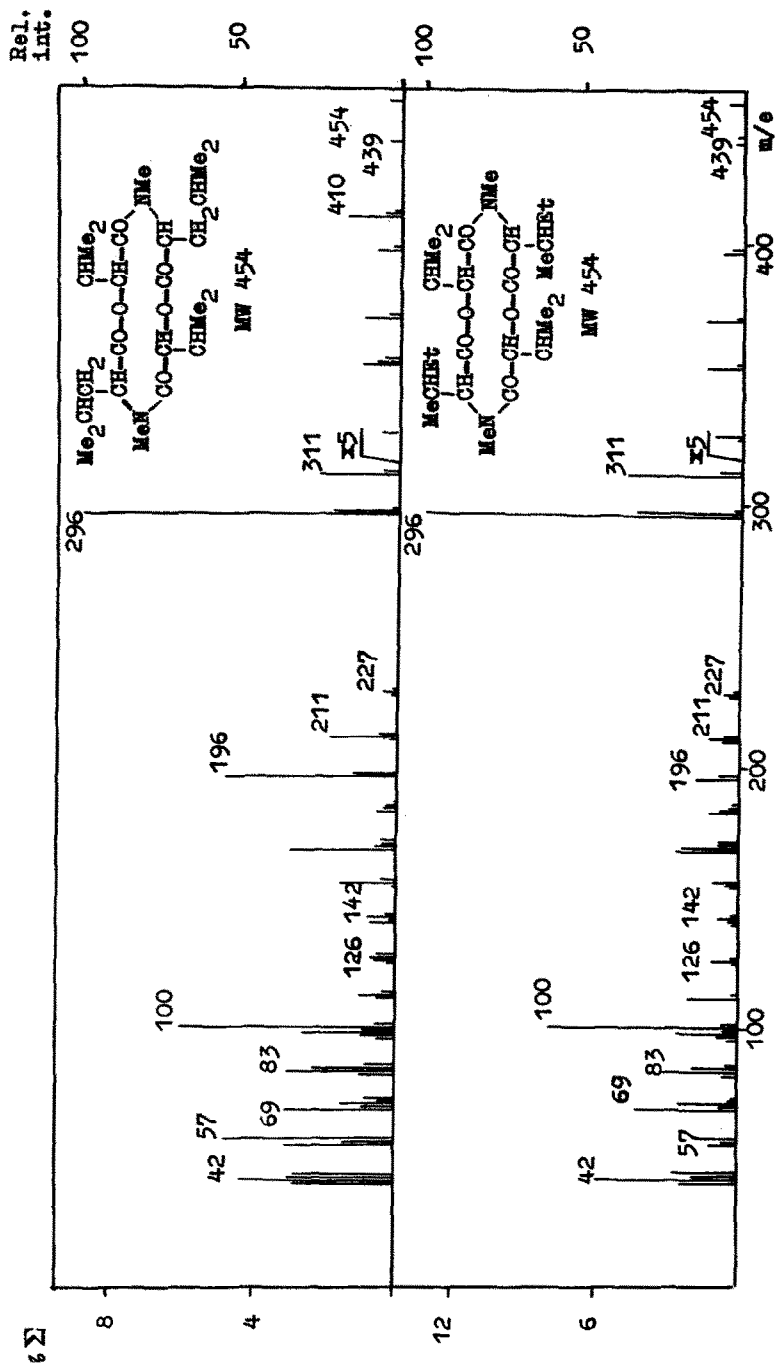
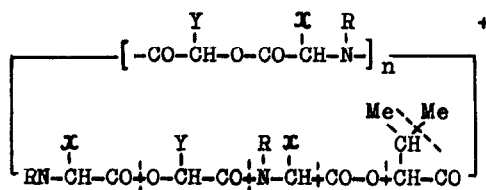


Fig. 2. Mass spectra of symmetrical cyclotetradepsipeptides.

being determined by the structure of the amino and hydroxy acid residues. The effect of the amino acid structure on the direction of the fragmentation is very strikingly displayed on comparing the intensities of the peaks  $F_2$ ,  $F_3$ ,  $F_4$  and  $F_6$  in the mass spectra of the isomeric 2,5-dioxomorpholines III and IV (Table 1).

The  $CO_2$  type of fragmentation (Scheme 2) is illustrated

Scheme 2  
 $CO_2$  type of cyclodepsipeptide fragmentation



$n = 0, 1, 2$ ;  $R = H, Me$ ;  $X = CHMe_2, CH_2CHMe_2, CH(Me)Et$ ;  $Y = Me, CHMe_2$

Table 2  
 $CO_2$  type of cyclotetradepsipeptide fragmentation

				Characteristic peaks m/e				
R	X	X'	$M^+$	$F_8$	$F_9$	$F_{10}$	$F_{11}$	$F_{12}$
V	H	$CHMe_2$	398	354	283	268	183	168
VI	Me	$CHMe_2$	426	382	297	282	197	182
VII	Me	$CH_2CHMe_2$	454	410	311	296	211	196
VIII	Me	$CH(Me)Et$	454	410	311	296	211	196
IX	Me	$CH_2CHMe_2$	454	410	311	296	211	196
X	Me	$CHMe_2$	440	396	311	296	211	196
					297	282	197	182
XI	Me	$CH(Me)Et$	440	396	311	296	211	196
					297	282	197	182

by the mass spectra of the series of cyclotetradepsipeptides (V-XI)(Table 2), possessing a rigid 12-membered ring and differing in the radicals of the amino acid residues, as well as in the substituents at the nitrogen atoms. After elimination of  $\text{CO}_2$  the resultant fragment  $F_8$  (M-44) evolves the grouping  $-\text{NR}-\text{CHX}$  (or  $\text{X}'$ )- belonging to one of the amino acid residues. This results in the formation of the fragment  $F_9$  which is further stabilized by loss of a methyl group leading to the appearance of the fragment  $F_{10}$ . In fact, in the case of symmetric cyclotetradepsipeptides V-VIII the mass spectra show the two strong peaks differing by 15 m.u., whereas with nonsymmetric cyclotetradepsipeptides X and XI (where the  $\text{X}'$  substituent differs from  $\text{X}$  by a homologous difference) the high mass region exhibits two pairs of strong peaks, the centers of which are 14 m.u. apart, the peaks within each pair being separated by 15 m.u., owing to stabilization brought about by loss of the methyl group (Figs. 1,2 and Table 2). Each of the characteristic fragments  $F_9$  and  $F_{10}$  may further eliminate a hydroxy acid residue ( $-\text{OCH}_2\text{CO}-$ ). Therefore with compounds V-XI (where  $\text{Y} = \text{CHMe}_2$ ) there are one or two pairs of peaks  $F_{11}$  and  $F_{12}$  differing by 100 m.u. from the corresponding characteristic peaks  $F_9$  and  $F_{10}$  respectively (Figs. 1,2 and Table 2).

The third type of fragmentation (Scheme 3) is characterized by formation of an acylaminoketene ion  $F_{13}$  as the result of the rupture of one ester bond and one ether bond. The rupture may be accompanied by rearrangements similar to those observed by Biemann (2) in the mass spectra of esters, amides and nitriles of aliphatic acids, as well as of linear tripeptides. Further decomposition of  $F_{13}$ , for example, with  $m/e$  184 for the cyclo-

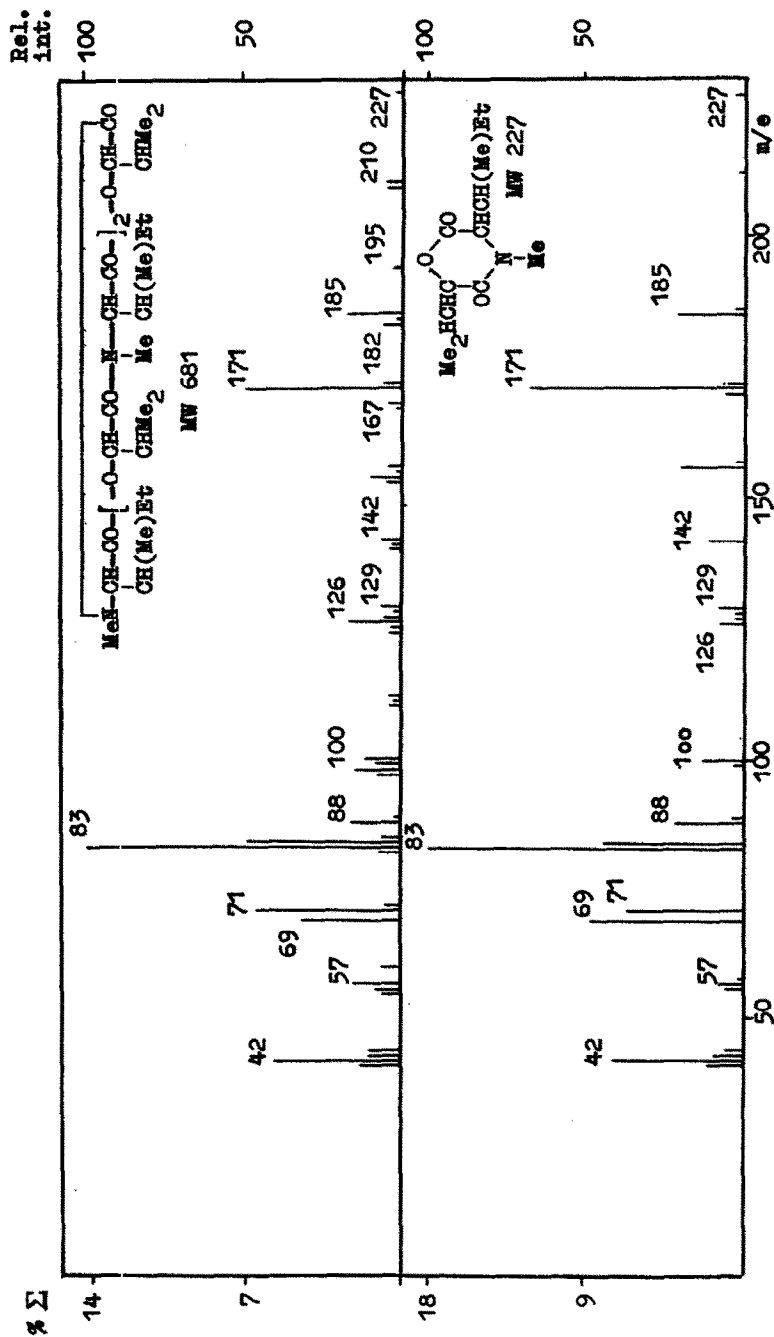


Fig. 3. Mass spectra of cyclodepsipeptide (low masses) and 2,5-dioxomorpholine.



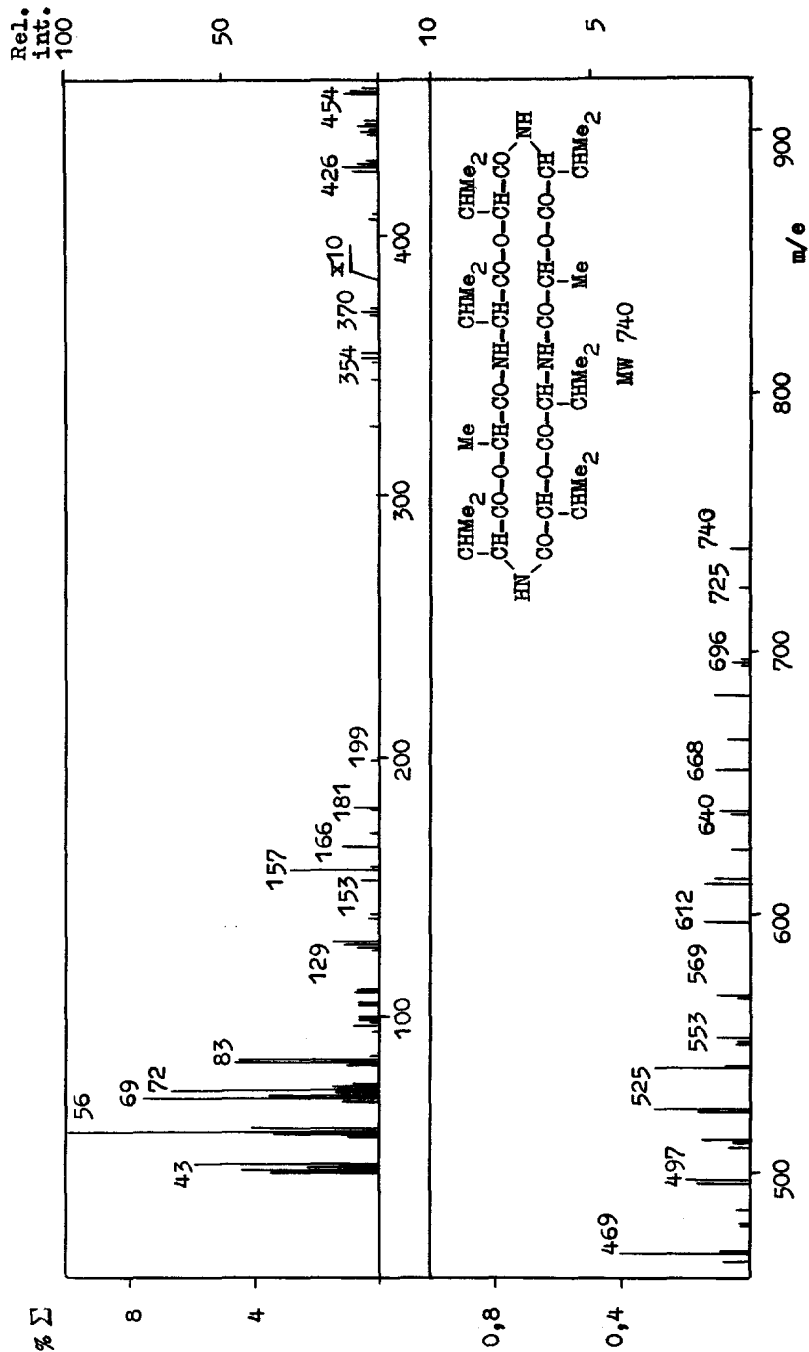
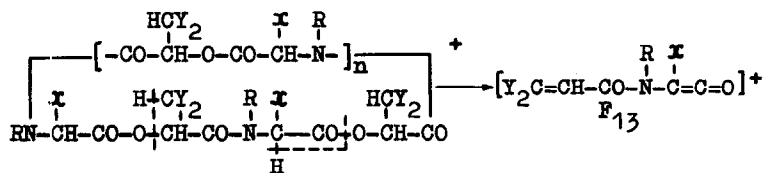


Fig. 4. Mass spectra of cyclooctadepsipeptide.

octadepsipeptide (Fig. 4) is associated with elimination of a Me and CO group, as a result of which the peak with  $m/e$  181 shifts by 15, 28 and 43 m.u. (peaks with  $m/e$  166, 153 and 138).

### Scheme 3

Acylaminoketene type of cyclodepsipeptide fragmentation



$n = 1, 2$ ;  $R = H, Me$ ;  $X = CHMe_2, CH_2CHMe_2, CH(Me)Et$ ;  $Y = H, Me$ .

The correctness of the sequences of fragment formation shown in Schemes 1-3 is confirmed by the presence of the corresponding metastable peaks in the mass spectra of the compounds investigated.

It thus follows that mass spectrometry of cyclodepsipeptides permits not only determination of the molecular weights of these compounds and their ring size but also elucidation of the nature of their amino and hydroxy acid components.

All the mass spectra were obtained at 150-225° and ionization potential 25-40v on the commercial mass spectrometer MX-1303 with a 90° analyzer and a stainless steel inlet system.

### REFERENCES

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